Mersana

Advantages of Polyacetal Polymer-Based Antibody Drug Conjugates Employing Cysteine Bioconjugation

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Summary

The application of polymers to antibody drug conjugates (ADC) can provide numerous advantages, including 1) significantly higher capacity for drug payload without sacrificing PK or tolerability; 2) utilization or alternative payloads not suitable for direct conjugation approaches; 3) improvement of physiocchemical properties of resulting ADCs; and 4) utilization of protein recognition scaffolds beyond the commonly used IgSs.

Herein we present results of a novel, biodegradable polyacetal polymer-based conjugation system to create next-generation ADCs. The basis of this new conjugation system is a hydrophilic, fully biodegradable polyacetal carrier (PHF or poly(1-hydroxymethylethylene hydroxymethyltormal, or Flexime®) modified with chemically orthogonal linkers. One linker is used to covalently attach a targeting moviely (mAb or alternative) varies of conjugation, while a second, chemically distinct linker is used to attach a drug payload and to control the mechanism and rate of drug release.

Previously we have reported highly efficacious polyacetal ADCs prepared by bioconjugation of the polymer to random yisine residues; in this report we present an alternative cysteinebased bioconjugation strategy. It is known that direct drug-cysteine linked ADCs result in destabilization of the protein, as the conjugation process necessarily disrupts inter-chain disulfide bridges. In contrast, the Fleximer conjugation approach via cysteines in the antibody hinge region allows for the formation of inter-chain bridge structures involving the polymer backbone, which provide restabilization of the overall construct, as evidenced by analytical methods such as SDS-PAGE and HPLC.

To demonstrate the benefits of this approach, we prepared Her-2 targeted ADCs with protein recognition scatfolds ranging in size from 15 KDa to 150 KDa, all targeting the Her-2 antigen, and bearing a proprietary Dolastiant derivative, XMT-1267, coupled to a Fleximer scatfold (Dolaflexin¹¹). These Dolaflexin ADCs were highly active and selective *in vitro* in Her-2 expressing cell lines. Furthermore, trastuzumab-Dolaflexin ADCs tested *in vivo* exhibited prolonged plasma exposure and tumor specific accumulation in the Her-2 expressing BT474 mouse xenograft model. The ADC was well-tolerated, and resulted in 100% tumor-free survivors at doces as low as 2 mg/kg.













Representative IHC of BT474 tumors 48 hours post administration
brown color – anti-CD31, blue color – anti-tuman IgG1 k





Dolaflexin ADC Plasma PK (SCID Mice, n=3)



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Discussion and Conclusions

 We have demonstrated that conjugation of the Fleximer-XMT-1267 (Dolaflexin) based cytotoxic drug conjugated to cysteine residues in the hinge region of the antibody results in highly reproducible, well-characterized ADCs which do not show the destabilization of the protein characteristic of conventional, direct cysteine-drug ADCs.

- Trastuzumab-Dolaflexin ADCs with a DAR of 20 are highly potent and selective in vitor in a variety of cell lines, consistent with antigen-dependent binding and internalization. Similarity, alternative Her-2 targeting moletles including at trastuzumab Fab fragment or a Her-2 targeted Affbody can be conjugated via cysteine residues to Dolaflexin, resulting in highly active and selective Her-2 targeted drug conjugates.
- In *in vivo* studies, the trastuzumab-Dolaflexin ADC with DAR of 20 was highly efficacious and displayed excellent pharmacokinetics, tolerability, tumor penetration, and target-specific tissue accumulation.

Additional examples of Dolaflexin-based ADCs employing antibodies other than trastuzumab will be reported in due course.

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